

Interaction between policosanol and prostacyclin on platelet aggregation in rats

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Palabras clave: policosanol, prostaciclina, interacción, agregación plaquetaria.
Key words: policosanol, prostacyclin, interaction, platelet aggregation.

RESUMEN. El policosanol es una mezcla de alcoholes alifáticos de elevado peso molecular, aislada y purificada de la cera de la caña de azúcar (*Saccharum officinarum*, L.), cuyo principal componente es el octacosanol. El tratamiento oral con policosanol reduce las concentraciones séricas de colesterol en diferentes modelos experimentales, voluntarios sanos y pacientes con hiperlipoproteíemia tipo II. Resultados anteriores han demostrado que la administración oral de policosanol inhibe la agregación plaquetaria inducida *in vivo* por colágeno y *ex vivo* por ADP en ratas. También ha sido demostrado una reducción de las concentraciones séricas de tromboxano B₂ en ratas, Mongolian gerbils y ratones, así como un incremento de la prostaciclina en suero de Mongolian gerbils en animales tratados con policosanol. Estos efectos también han sido observados en voluntarios sanos y pacientes con hipercolesterolemia tipo II. La prostaciclina es el más potente inhibidor de la agregación plaquetaria descubierto hasta el momento. El objetivo de este estudio ha sido investigar los efectos de la interacción del policosanol y la prostaciclina sobre la agregación plaquetaria *ex vivo* en ratas. La sensibilidad del plasma rico en plaquetas (PRP) a la prostaciclina exógena fue evaluada midiendo el porcentaje de inhibición de la agregación inducida por ADP (pre-incubado con prostaciclina) y el porcentaje de desagregación después de la adición de prostaciclina cuando la agregación máxima e irreversible al ADP fue alcanzada. Los resultados de este estudio muestran que no solo, como ha sido reportado en trabajos anteriores, la agregación inducida por ADP en PRP de animales pre-tratados con policosanol (200 mg/kg) fue significativamente menor que en animales controles sino, que la sensibilidad a la prostaciclina exógena fue mayor en el PRP de animales tratados con policosanol.

ABSTRACT. Policosanol is a mixture of higher molecular aliphatic primary alcohols, isolated from sugar cane wax, (*Saccharum officinarum*, L.) whose main component is octacosanol. Oral treatment with policosanol reduced serum cholesterol levels in different experimental models, healthy volunteers and patients with type II hyperliproteinemia. Previous results showed that policosanol orally administered inhibits platelet aggregation induced *in vivo* by collagen and induced *ex vivo* by ADP in rats as well as significantly reduces serum thromboxane B₂ levels in rats, Mongolian gerbils and mice and increases prostacyclin serum levels in Mongolian gerbils. It has been shown that policosanol also exhibits antiplatelet properties in human beings. Prostacyclin (Pgl₂) is the most potent endogenous inhibitor of platelet aggregation discovered so far. This study was undertaken to investigate the effect of the interaction of policosanol and Pgl₂ on *ex vivo* platelet aggregation in rats. The sensitivity of PRP to exogenous Pgl₂ was tested measuring the percentage of inhibition on ADP-induced aggregation (pre-incubation with Pgl₂) and the percentage of disaggregation after addition of Pgl₂ when the maximal and irreversible response of ADP was reached. The results showed that not only, as has been previously reported, ADP induced aggregation in platelet rich plasma (PRP) of policosanol (200 mg/kg) pretreated animals was significantly lower than in control animals but also that the sensitivity to exogenous Pgl₂ of PRP in policosanol pretreated animals was also higher than in the controls.

INTRODUCTION

Policosanol is a mixture of higher molecular aliphatic primary alcohols, isolated from sugar cane wax, (*Saccharum officinarum*, L.) whose main component is octacosanol, followed by triacontanol and hexacosanol, while the other alcohols (tetracosanol, heptacosanol, nonacosanol, dotriacontanol and tetratriacontanol) are minor components.

Oral treatment with policosanol reduced serum cholesterol levels in different experimental models,^{1,2} healthy volunteers³ and patients with type II hyperliproteinemia.⁴⁻⁹

Previous results showed that policosanol orally administered inhibits platelet aggregation induced *in vivo* by collagen and induced *ex vivo* by ADP in rats¹⁰ as well as significantly reduces serum thromboxane B₂ levels in rats, Mongolian gerbils and mice¹⁰⁻¹² and increases prostacyclin serum levels in Mongolian gerbils.¹¹

It has been shown that policosanol also exhibits antiplatelet properties in human beings.¹³

Prostacyclin (Pgl₂) is the most potent endogenous inhibitor of platelet aggregation discovered so far. It also disaggregates platelets *in vitro*¹⁴⁻¹⁵ and can prevent or reverse *in vivo* or *ex vivo* platelet aggregation.¹⁶ An increase in cAMP levels of platelets has been proposed as the mechanism of this antiplatelet effect of Pgl₂.¹⁷

There are alternative approaches for enhancing the therapeutic action of Pgl₂. One of them is the concomitant use of a phosphodiesterase in-

hibitor such as theophylline or dipyridamole which may potentiate the effect of Pgl_2 by preventing the breakdown of intracellular cAMP in platelets.¹⁵⁻¹⁸

Taking into account these facts this study was undertaken to investigate the effect of the interaction of policosanol and Pgl_2 on *ex vivo* platelet aggregation in rats.

MATERIAL AND METHODS

Animals

Male Sprague Dawley rats weighing 250-300 g obtained from Centro Nacional para la Producción de Animales de Laboratorio were used and adapted to experimental conditions for 7 d with food and water *ad libitum*. Rats were randomly distributed in two groups.

Reagents

ADP equine muscle (Sigma); acacia gum (BDH); prostacyclin (Sigma).

Administration and dosage

Policosanol was prepared as a suspension in acacia gum (10 mg/kg) and administered by gavage at 1 mL/200 g body weight. Control animals received the same volume of acacia gum vehicle. All animals were fasted but had free access to water for 16 h prior to the experiment.

Pgl_2 was kept in a stock solution (1 mmol/L ethanol) and diluted in Tris HCl buffer pH 7.4 immediately before use.

Platelet aggregation induction

Two hours after treatments all animals were anaesthetized with 40 mg/kg sodium pentobarbital and blood samples were drawn from cava vein and mixed with 3.8 % sodium citrate (9 volumes of blood per 1 of anticoagulant). Blood was centrifuged at 250 g for 10 min to obtain platelet-rich-plasma (PRP). Once PRP was isolated, the remainder was centrifuged at 1 300 g for 15 min to obtain platelet-poor-plasma (PPP).

Platelet aggregation was quantified by Born turbidimetric method.¹⁹ PRP (250 μL) aliquots were preincubated for 2 min at 37 °C on a Elvi 840 aggregometer, stirred at 1 000 r/min and stimulated by the addition of 5 or 12 $\mu\text{mol/L}$ ADP (concentrations obtained in the incubation medium). Platelet aggregation levels were measured after calibrating the equipment at 0 % light transmission for PRP and at 100 % for PPP. Aggregation curves were recorded for 5 min.

The antiaggregatory activity of Pgl_2 was assessed by determining the percentage of inhibition on ADP (5 $\mu\text{mol/L}$)-induced PRP aggregation in the presence of Pgl_2 ($1 \cdot 10^{-8}$ mol/L) or Tris HCl buffer added to PRP 1 min before ADP in PRP of controls and policosanol pretreated animals.

The disaggregatory activity of Pgl_2 was assessed by determining the percentage of disaggregation on ADP (12 $\mu\text{mol/L}$)-induced PRP irreversible aggregation after the addition of Pgl_2 ($2 \cdot 10^{-8}$ mol/L).

Statistical analysis

Comparisons between groups were done using the Mann-Whitney U test.

RESULTS

The results showed that not only, as has been previously reported, ADP induced aggregation in PRP of policosanol (200 mg/kg) pretreated animals was significantly lower than in control animals but also that the sensitivity to exogenous Pgl_2 of PRP in policosanol pretreated animals was also higher than in the control's (Table 1).

The sensitivity of PRP to exogenous Pgl_2 was tested measuring the percentage of inhibition on ADP-induced aggregation (pre-incubation with Pgl_2) and the percentage of disaggregation after addition of Pgl_2 when the maximal and irreversible response of ADP was reached (Table 2).

DISCUSSION

A number of disease states have now been related to an imbalance in the vascular Pgl_2 and thromboxane A_2 (TxA_2) system.

Patients with arterial thrombosis, deep vein thrombosis and recurrent venous thrombosis produce more of the proaggregatory compounds: PG endoperoxides and TxA_2 in their platelets. Atherosclerotic rabbits and patients who have survived myocardial infarction produce more TxA_2 than controls and they are also very sensitive to other aggregating agents.²⁰⁻²¹

Not only diabetic rats release more TxA_2 than normal rats but also their blood vessels show reduced generation of Pgl_2 ,²² blood vessels from diabetic patients show reduced production of Pgl_2 ,^{23, 24} and circulating levels of 6 oxo Pgf1a , the metabolite of Pgl_2 are reduced in diabetic patients with proliferative retinopathy.²⁵

Smooth muscle cells obtained from atherosclerotic lesions and cultured *in vitro* also produced less Pgl_2 than normal vascular smooth muscle.²⁶

Increased levels of low density proteins (LDL) has been suggested to be a risk factor associated with ischaemic heart disease,^{27,28} meanwhile an inverse relation is shown regarding high density lipoprotein (HDL) levels. LDL reduces the generation of prostacyclin-like substance from human endothelial cells whereas HDL stimulated Pgl_2 synthesis. It is likely that the lipid per-

Table 1. Sensitive to exogenous Pgl_2 in PRP of control and policosanol treated animals.

Treatment	n	Aggregation		Inhibition
		(Buffer)	(PgI ₂)	
		(%)		
Control	6	59.5 ± 18.6	38.6 ± 4.0	31.3 ± 6.4
Policosanol 20 mg/kg	6	49.1 ± 20.3	23.5 ± 11.0	46.9 ± 10.0*
Policosanol 200 mg/kg	6	31.2 ± 14.6*	10.6 ± 9.0**	71.0 ± 10.6**

*p < 0.05; **p < 0.01. Mann Whitney U test. Comparisons with control group.

Table 2. Disaggregation induced by Pgl_2 in control and policosanol treated animals.

Treatment	n	Disaggregation
(%)		
Control	6	15.4 $\pm$ 11.9
Policosanol 20 mg/kg	6	58.8 $\pm$ 3.6*
Policosanol 200 mg/kg	6	61.6 $\pm$ 15.5*

*p < 0.05 Mann Whitney U test.

oxides found in LDL (and not in HDL) may be responsible for the inhibition of Pgl_2 synthesis.²⁹

Blood vessels of the brain of all animals species, including man, generate Pgl_2 .^{30,31} Several investigators have suggested that Pgl_2 may be involved in the regulation of blood flow in the brain.³²

These results suggest that prostanoids (TxB_2 and Pgl_2) have a role in platelet function and are involved in cardiovascular as well as cerebrovascular disorders.

This concept has provided the basis for the development of compounds with therapeutic potential in reducing intravascular thrombus formation as: cyclooxygenase inhibitors, thromboxane synthetase inhibitors, thromboxane antagonists and Pgl_2 analogues.

Previous results have shown that single doses of policosanol not only are able to reduce TxA_2 and malondyaldehyde,¹⁰ but also increases Pgl_2 serum levels in several animal models,¹¹ suggesting an thromboxane synthetase inhibitor like effect.

On the other hand, platelets of hyperlipoproteinemic (HLP) patients exhibit a significantly reduced sensitivity to prostacyclin,³³ iloprost and PGE_1 .³⁴ Thus, considerably higher concentrations of these prostaglandins are required to inhibit platelet function in HLP than in healthy controls.

Oliva *et al.*³⁴ have shown that the number of prostacyclin binding sites in platelet membranes, prepared from platelet-rich plasma, was significantly reduced in type IIa HLP whereas the K_D , indicating the affinity of the binding sites, remained unchanged.

Jaschonek *et al.*³⁵ using cholesterol-hemisuccinate-loaded platelets, have shown that incorporation of cholesterol results in a decrease of membrane fluidity and an apparent loss of iloprost binding sites while administration of local anesthetics, leading to increased membrane lipid fluidity, exerted the opposite effect.

It has been argued that if the changes in platelet reactivity, sensitivity to Pgl_2 , and eicosanoids biosynthesis reported in hypercholesterolemia are a direct consequence of elevated plasma lipid levels, then one would expect that cholesterol lowering by dietary means, plasmapheresis, or lipid-lowering drugs would result in normalization of platelet function.³⁶ In fact, LDL apheresis, fibric acid derivatives, ion exchange resins, probucol, and

HMG-CoA reductase inhibitors have all been evaluated for their effects on platelet function. In these studies, except those carried out with simvastatin, despite LDL-C reductions on the order of 10 to 30 %, no change in platelet aggregation³⁷ and platelet sensitivity to iloprost³⁸ could be demonstrated *ex vivo*.

The results of the present work show that policosanol at the same doses reduces thromboxane and increases Pgl_2 serum levels, enhances exogenous Pgl_2 antiaggregatory effect in rats.

The fact that policosanol increases not only Pgl_2 serum levels but also the sensitivity of exogenous Pgl_2 suggests that it could act preventing the breakdown of Pgl_2 to its inactive metabolite (6 oxo $\text{Pgf}_{1\alpha}$).

Although the exact mechanism of antiplatelet action of policosanol is unknown, these results can explain at least partially the protective effect of policosanol on atherosclerotic lesions in experimental models³⁹ and human beings.⁴⁰

BIBLIOGRAPHY

1. Arruzazabala M.L., Carbajal D., Más R., Valdés S., and Laguna A. Cholesterol-lowering effects of policosanol in rabbits. *Biological Research*, 27, 205, 1994.
2. Rodríguez-Echenique C., Mesa R., Más R., y Castaño G. Estudio del efecto sobre lípidos y lipoproteínas séricas y de la tolerancia al tratamiento oral con dosis crecientes de policosanol en monos *Macaca arctoides*. *Arch. Venezolanas de Farmacología y Terapéutica*, 11, 74, 1992.
3. Hernández F., Illnait J., Más R., Castaño G., Fernández L., González M., Cordovi N. and Fernández J. Efficacy and safety of policosanol in patients with primary hypercholesterolemia. *Current Therapeutic. Research*, 51, 568, 1992.
4. Pons P., Más R., Illnait J., Fernández L., Rodríguez M., Robaina C., and Fernández J. Efficacy and safety of policosanol in patients with primary hypercholesterolemia. *Current Therapeutic. Research*, 52, 507, 1992.
5. Pons P., Jiménez A., Rodríguez M., Illnait J., Más R., Fernández L., and Fernández J. Effects of policosanol in elderly hypercholesterolemic Patients. *Current Therapeutic. Research*, 53, 265, 1993.
6. Illnait J., Castaño G., Nodarse N., Pontigas V., Fernández L. and más R. Efectos del atermixol (PPG) sobre la hiperlipoproteinemia del tipo II. *Rev CENIC Ciencias Biológicas*, 22, 74, 1991.
7. Castaño G., Zardoya R., Illnait J., Más R., Fernández L., Surribas E., Nodarse M., Fernández J. Efectos del tratamiento con atermixol (PPG) (5 mg) en pacientes con hiperlipoproteinemia tipo II. *PCM*, 5, 21, 1991.

8. Soltero I., Fuenmayor I., Colmenares J. Estudio comparativo doble ciego de la eficacia y tolerancia del policosanol vs. bezafibrato en pacientes con hiperlipoproteinemia tipo II. *Arch. Venezolanas Farmac y Terap*, 12, 71, 1993.
9. Pons P., Rodríguez M., Robaina C., Illnait J., Más R., Fernández L., Fernández J. Effects of policosanol successive dose increases on lipid profile of patients with type II hypercholesterolemia and tolerability to treatment. *Clin. Pharmacol. Res.*, XIV, 27, 1994.
10. Arruzazabala M.L.; Carbajal D.; Más R.; García M.; and Fraga V. Effects of policosanol on platelet aggregation in rats. *Thrombosis Res.*, 69, 321, 1993.
11. Arruzazabala M.L., Molina V., Carbajal D., Valdés S., and Más R. Effect of policosanol on cerebral ischaemia in Mongolian gerbils: Role of prostacyclin and thromboxane A2. *Prost. Leuk. Essent Fatty Acids*, 49, 695, 1993a.
12. Arruzazabala M.L., Carbajal D., Molina V., Valdés S., Más R., García M. Estudio farmacológico de la interacción entre el policosanol y la aspirina en animales de experimentación. *Rev. Iberoamer. Tromb. Hemost.*, 5, 17, 1992.
13. Arruzazabala M.L., Valdés S., Más R., Fernández L. and Carbajal D. Effect of policosanol successive dose increases on platelet aggregation in healthy volunteers. *Pharmacol. Res.*, 33, 131, 1996.
14. Moncada S., Gryglewski R., Bunting S., Vane J.R. An enzyme isolated from arteries transforms prostaglandin endoperoxides to an unstable substance that inhibits platelet aggregation. *Nature*, 263, 663, 1976.
15. Ubatuba F.B., Moncada S., Vane J.R. The effect of prostacyclin (Pgl_2) on platelet behaviour. Thrombus formation *in vivo* and bleeding time. *Thromb. Res.*, 41, 425, 1979.
16. Karim S.M.M., Adaikan P.G. Some pharmacological studies with prostacyclin in baboon and man. In: Vane J.R. Bergstrom S ed. Prostacyclin, New York: Raven Press. 419, 1979.
17. Gorman R.R., Bunting S., Vane J.R. Effects of prostacyclin (Pgl_2) and the effect of phosphodiesterase inhibitors on platelet aggregation. *Pharmacological Res. Comm.*, 11, 605, 1979.
18. Jorgensen K.A., Dyerberg J., Stoffersen E. Prostacyclin (Pgl_2) and the effect of phosphodiesterase inhibitors on platelet aggregation. *Pharmacological Res. Comm.*, 11, 605, 1979.
19. Born G. Aggregation of blood platelets by adenosine diphosphate and its reversal. *Nature (London)*, 194, 927, 1962.
20. Shimamoto T., Kobayashi M., Takahashi T., Takashima Y., Sakamoto M., Morooka S. An observation of thromboxane A_2 in arterial blood after cholesterol feeding in rabbits. *Japan Heart Journal*, 19, 748, 1978.

21. Szezeklik A., Gryglewski R.J., Musial J., Grodzinka L., Serwonska M., Marcinkiewicz E. Thromboxane generation and platelet aggregation in survivals of myocardial infarction. **Thromb and Haemostasis**, **40**, 66, 1978.
22. Harrison H.E., Reece A.H., Johnson M. Decreased vascular prostacyclin in experimental diabetes. **Life Sciences**, **23**, 351, 1978.
23. Johnson M., Harrison H.E., Raftery A.T., Elder J.B. Vascular prostacyclin may be reduced in diabetes in man. **Lancet**, **1**, 325, 1979.
24. Davies T.M.E., Brown E., Finch D.R., Mitchell M.D., Turner R.C. *In vitro* venous prostacyclin production plasma 6 keto prostaglandin F1a concentrations and diabetic retinopathy. **Medical Journal**, **282**, 1259, 1981.
25. Dollery C.T., Friedman L.A., Hensby C.N., Kohner E., Lewis P.J., Porta M., Webster J. Circulating prostacyclin may be reduced in diabetes. **Lancet**, **2**, 1365, 1979.
26. Larrue J., Rigaud M., Daret D., Demond J., Durand J., Brincaud H. Prostacyclin production by cultured smooth muscle cells from atherosclerotic rabbit aorta. **Nature**, **285**, 480, 1980.
27. Streja D., Steiner G., Kwiterovich P.O. Plasma high-density lipoproteins and ischemic heart disease. **Annals of Internal Medicine**, **89**, 871, 1978.
28. Kannel W.B., Castelli W.P., Gordon T. Cholesterol in the prediction of atherosclerotic diseases. **Annals of Internal Medicine**, **90**, 85, 1979.
29. Gryglewski R.J., Szezeklik A. Prostacyclin and atherosclerosis. In: Lewis P.J., O'Grady J. Ed. **Clinical Pharmacology of Prostacyclin**. New York, Raven Press, 89, 1981.
30. Boullin D.J., Bunting S., Blaso W.P., Hunt T.M., Moncada S. Responses of human and baboon arteries to prostaglandin endoperoxides and biologically generated and synthetic prostacyclin: their relevance to cerebral arterial spasm in man. **British Journal of Clinical Pharmacology**, **7**, 139, 1979.
31. Abdel-Halim M.S., Horst von H., Meyerson B., Sachs C., Anggard E. Prostaglandin profile in tissue and blood vessels from human brain. **Journal of Neurochemistry**, **34**, 1331, 1980.
32. Pickard J.D. Role of prostaglandins and arachidonic acid derivatives in the coupling of cerebral blood flow to cerebral metabolism. **Journal of cerebral Blood Flow and Metabolism**, **1**, 361, 1981.
33. Colli S., Maderna P., Tremoli E., Baraldi A., Rovati G., Gianfranceschi G., Nicosia S. Prostacyclin-lipoprotein interactions. Studies on human platelet aggregation and adenylate cyclase. **Biochem. Pharmacol.**, **34**: 2451, 1993.
34. Oliva D.W., Maderna P., Accomazzo M.R., Nicosia S., Tremoli E. Iloprost binding and inhibition of aggregation in platelet rich plasma. Differences between normal and type IIa hypercholesterolemic subjects. **Biochem. Pharmacol.**, **38**, 39, 1989.
35. Jaschonek K., Funck M., Muller C.P. Modulation of platelet 3H iloprost binding by cholesterol, dibucaine and pentoxifylline. In: Prostaglandins in clinical research. Schror, K. Sinzinger H. (Ed.). Alan R. Liss, New York, 312-325, 1989.
36. Tremoli E., Colli S., Maderna P., Baldassarre D., di Mino G. Hypercholesterolemia and platelets. **Semin Thromb. Hemost.**, **19**, 115, 1993.
37. Schror K. Platelet reactivity and arachidonic acid metabolism in type II hyperlipoproteinemia and its modification by cholesterol-lowering agents. **Eicosanoids**, **3**, 67, 1990.
38. Notarbartolo A., Davi G., Aversa M., Barbagallo C., Ganci A., Giammarresi C., La Placa F., Patrono C. Inhibition of thromboxane biosynthesis and platelet function by simvastatin in type IIa Hypercholesterolemia. **Atherosclerosis, Thrombosis and vascular Biology**, **15**, 247, 1995.
39. Noa M., Más R., De la Rosa M.C., Magraner J. Effect of policosanol on lipofundin-induced lesions in rats. **J. Pharmacy Pharmacol.**, **47**, 289, 1995.
40. Batista J., Stuser R., Penichet M. and Uget E. Doppler-Ultrasound pilot study of the effects of long-term policosanol therapy on carotid-vertebral atherosclerosis. **Curr. Ther. Res.**, **56**, 906, 1995.



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